

Synthesis and NMR spectral study of some 5-aryl-3-methylcyclohex-2-enones and 5*r*-aryl-3*t*-cyano-3*c*-methylcyclohexanones

S Kamatchi, R T Sabapathy Mohan, R Gomathi & K Pandiarajan*

Department of Chemistry, Annamalai University, Annamalainagar 608 002, India

E-mail: profkprajan@yahoo.co.in

Received 13 May 2008; accepted (revised) 13 January 2009

Eight 5-aryl-3-methylcyclohex-2-enones **2a-h** (Ar = C₆H₅, *p*-ClC₆H₄, *p*-MeOC₆H₄, *p*-O₂NC₆H₄, *p*-MeC₆H₄, *p*-FC₆H₄, *p*-Me₂NC₆H₄ or *m*-O₂NC₆H₄) and seven 5*r*-aryl-3*t*-cyano-3*c*-methylcyclohexanones **3a-c** and **3e-h** (Ar = C₆H₅, *p*-ClC₆H₄, *p*-MeOC₆H₄, *p*-MeC₆H₄, *p*-FC₆H₄, *p*-Me₂NC₆H₄ or *m*-O₂NC₆H₄) have been synthesized and characterised by ¹H and ¹³C NMR spectra. For **2a** (Ar = C₆H₅) SEFT and HMBC spectra have been recorded. For **2e** (Ar = *p*-MeC₆H₄) HSQC and HMBC spectra have been recorded. For **3a** (Ar = C₆H₅) HOMOCOSY, NOESY, HSQC and SEFT spectra have been recorded. Analysis of these spectra suggests that cyclohex-2-enones adopt sofa conformation with one of the methylene protons at C-4 and C-6 *anti* to H-5 and the other methylene proton *gauche* to H-5. The effect of the phenyl and methyl groups on the chemical shifts of protons and carbons in cyclohexenone ring are discussed. The cyanocyclohexanones adopt chair conformation with axial orientation of cyano group and equatorial orientations of the methyl and phenyl groups. The probable values for the proton chemical shifts of the various protons in the cyclohexanone ring of *cis*-3-methyl-5-phenylcyclohexanone **6** have been computed from the observed proton chemical shifts of **3a** and the known effects of axial cyano group. The possible values for the ¹³C chemical shifts of the cyclohexanone ring carbons in **6** have been computed from the ¹³C chemical shifts of cyclohexanone and known effects of phenyl and methyl groups. Comparison of these values with the chemical shifts of protons and carbons in **2a** suggests that the double bond at C-2 influences the ¹H chemical shifts by polarization, magnetic anisotropy and electronegativity effects and the ¹³C chemical shifts by polarization and electronegativity effects.

Keywords: Cyclohex-2-enones, cyanocyclohexanones, ¹H NMR, ¹³C NMR, conformation

Cyclohex-2-enones are useful in the synthesis of organic compounds¹⁻⁴. Attempts to prepare β-cyanoketones by the addition of hydrocyanic acid to α,β-unsaturated ketones have resulted in the formation of mixture of products⁵⁻⁷. However, β-cyanoketones are formed exclusively when α,β-unsaturated ketones are treated with a mixture of NH₄Cl and KCN⁸. Though a number of NMR spectral studies⁹⁻¹⁷ have been made on saturated six-membered compounds with and without heteroatoms in the ring, to our knowledge, only one NMR spectral study¹⁸ has been reported on cyclohexenones. Even in this study the chemical shifts and coupling constants for the protons on saturated carbons have not been determined precisely.

There are several NMR spectral studies^{10-12,14} on heterocyclic ketones, reporting the ¹H chemical shifts and proton-proton coupling constants precisely. However, such studies are rare on cyclohexanones. Recently, for the heavily substituted 2*r*,4*c*-biscarbalkoxy-3*t*-aryl-5*c*-hydroxy-5*t*-methylcyclohexanones **1** such data have been reported¹⁵⁻¹⁷. However, it is desirable to

investigate cyclohexanones with less number of substituents.

This paper reports the synthesis and NMR spectral study of eight 5-aryl-3-methylcyclohex-2-enones **2a-h** and seven 5*r*-aryl-3*t*-cyano-3*c*-methylcyclohexanones **3a**, **3b**, **3c**, **3e**, **3f**, **3g** and **3h**.

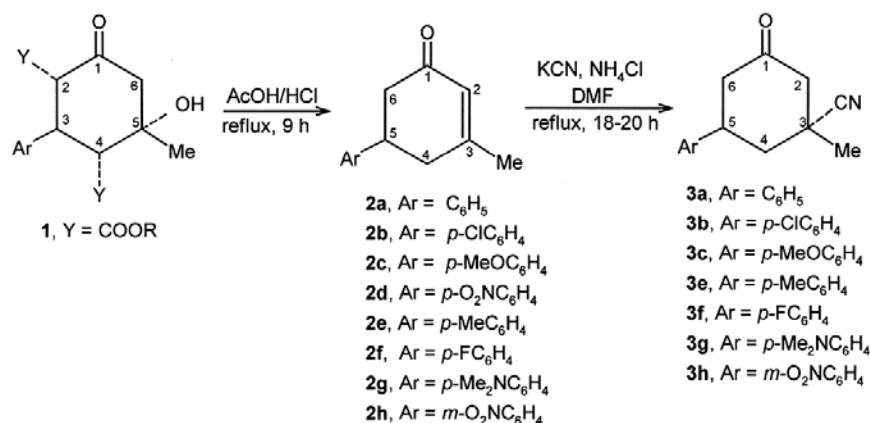
Results and Discussion

Synthesis and characterisation of compounds

Compounds included in the present study were obtained from **1** (Ref. 15-17), by reactions shown in **Scheme I**. The numbering of carbon atoms also is shown in **Scheme I**. All compounds were characterised by their IR spectra. Elemental analysis was done for **2b**, **2c**, **2d** and **3a**. Mass spectrum was recorded for **3a**. The physical data are given in **Table I** and the results of elemental analysis are given in **Table II**.

Cyclohex-2-enones **2a-h**

In the ¹H NMR spectrum of **2a** the aromatic protons appeared as a multiplet at δ 7.32. There was a



Scheme I

Table I—Physical data on cyclohexenones and cyanocyclohexanones

Ar	2			3		
	Yield %	m.p. °C	IR cm ⁻¹	Yield %	m.p. °C	IR cm ⁻¹
C ₆ H ₅	60	Liquid	1685	85	95	1715, 2231
<i>p</i> -ClC ₆ H ₄	72	55	1662	70	85	1710, 2235
<i>p</i> -MeOC ₆ H ₄	69	67	1659	70	61	1717, 2235
<i>p</i> -O ₂ NC ₆ H ₄	65	120	1659			
<i>p</i> -MeC ₆ H ₄	80	42	1667	80	103	1707, 2235
<i>p</i> -FC ₆ H ₄	70	45	1664	65	Liquid	1718, 2230
<i>p</i> -Me ₂ NC ₆ H ₄	65	47	1659	80	115	1711, 2235
<i>m</i> -O ₂ NC ₆ H ₄	75	105	1659	70	114	1722, 2238

Table II—Results of elemental analysis of typical compounds 2b-d and 3a

Compound	Experimental (%)			Calculated for the molecular formula (%)		
	C	H	N	C	H	N
2b	70.68	5.94		70.75	5.90	
2c	77.70	7.51		77.78	7.41	
2d	67.46	5.70	6.13	67.53	5.63	6.06
3a	78.89	6.99	6.82	78.87	7.04	6.57

singlet at δ 6.00 due to H-2. The benzylic proton H-5 appeared as a multiplet at δ 3.34. The methylene protons at C-4 and C-6 appeared as a overlap of two AB parts in the range 2.4-2.6 ppm. The methyl protons at C-3 appeared as a singlet at δ 2.03. The ¹³C NMR spectra of 2a-h were almost identical except for the aromatic part.

In the ¹³C NMR spectrum of 2a the carbonyl carbon appeared at δ 199.1. There were two weak signals at δ 161.7 and 143.3, which were assigned to C-3 and *ipso* carbon, respectively. Two strong signals at δ 127.0 and 128.8 were assigned to the *ortho* and

meta carbons of the phenyl group. There were two signals at δ 126.8 and 126.6 with almost half the intensity of the signals at δ 127.0 and 128.8. There were three signals at δ 43.9, 40.8 and 39.0 due to the saturated carbons of the cyclohexenone ring. The carbon of the methyl group at C-3 appeared at δ 24.4. From the SEFT spectrum of 2a it was found that the signals at δ 43.9 and 39.0 are due to methylene carbons. From the HMBC spectrum of 2a it was found that the signal at δ 39.0 is due to C-4 because this signal had a correlation with the methyl protons. From the other correlations in the HMBC spectrum of 2a and from the correlations in the HSQC and HMBC spectra of 2e the signals in the ¹H and ¹³C NMR spectra of cyclohex-2-enones 2a-h could be assigned unambiguously. The ¹H and ¹³C signals of the aromatic ring in 2b-h were assigned by known substituent effects^{19,20}. In 2b-h the signals due to C-2 was distinguished from the signals for the other aromatic carbons based on intensity.

From the HSQC spectrum of 2e it was found that the AB part due to the methylene protons at C-6 is at a higher frequency than that due to the methylene protons at C-4. The first six lines of this AB part

could be clearly seen. From the frequencies of these six lines the coupling constants $J_{5,6}$, $J_{5',6}$ and $J_{5,5'}$ were calculated using second-order analysis. The correct chemical shifts of the methylene protons H-6 and H-6' were also calculated. The frequencies of all lines in the signal for H-5 were given by the computers in the case of **2a**. The spacing between the first and last lines should correspond to the sum of $J_{5,6}$, $J_{5',6}$, $J_{4,5}$ and $J_{4',5}$. Hence, the sum of $J_{4,5}$ and $J_{4',5}$ could be obtained for **2a**. In all cases the $J_{5,6}$, $J_{5',6}$ and $J_{5,5'}$ could be determined. However, only in the cases of **2a**, **2b** and **2f** the sum of $J_{4,5}$ and $J_{4',5}$ could be determined. The proton chemical shifts and coupling constants of **2a-h**

are given in **Table III**. The ^{13}C chemical shifts are given in **Table IV**.

The chemical shifts of various protons in cyclohex-2-enone **4** (Ref. 21) and 3-methyl-cyclohex-2-enone **5** (Ref. 21) are shown in **Figure 1**. It is seen that in **4** the methylene protons at C-4, C-5 and C-6 give rise to average signals for two interchangeable environments. The appearance of the methylene protons at C-4 and C-6 in **2a-h** as two AB parts suggests that the aryl group at C-5 holds the cyclohex-2-enone ring in a fixed conformation.

Analysis of the *vicinal* coupling constants about the C(5)-C(6) bond, using the method of Slessor and

Table III — ^1H NMR spectral data of cyclohex-2-enones **2a-h**

Compound	Chemical shifts (δ , ppm) ^a							Coupling constant (J , Hz) ^b		
	H-2	H-4	H-5	H-6	H-6'	CH ₃	Aromatic ^c	$J_{5,6}$	$J_{5,6'}$	$J_{5,5'}$
2a	6.00	2.47	3.34	2.66	2.58	2.03	7.32	4.1	13.3	16.2
2b	6.03	2.55	3.36	2.67	2.57	2.07	7.38 (<i>o</i>) 7.22 (<i>m</i>)	4.1	13.5	16.1
2c ^d	6.06	2.58	3.35	2.71	2.61	2.09	7.35 (<i>o</i>) 6.96 (<i>m</i>)	4.0	13.6	16.1
2d	6.02	2.55	3.47	2.67	2.58	2.04	7.42 (<i>o</i>) 8.23 (<i>m</i>)	4.0	13.3	16.3
2e ^e	5.99	2.54	3.30	2.64	2.56	2.03	7.20 (<i>o</i>) 7.16 (<i>m</i>)	3.9	13.7	16.4
2f	6.01	2.54	3.34	2.66	2.56	2.05	7.25 (<i>o</i>) 7.06 (<i>m</i>)	4.0	13.5	16.2
2g ^f	6.00	2.53	3.27	2.67	2.57	2.04	7.35 (<i>o</i>) 6.77 (<i>m</i>)	3.8	13.6	16.2
2h	6.00	2.54	3.44	2.67	2.56	2.02	8.12, 7.56 (<i>o</i>) 7.51 (<i>m</i>) 7.56 (<i>p</i>)	3.5	13.6	16.1

^a For the methylene protons at C-4 the reported chemical shift corresponds to the centre of the AB part.

^b The values of ($J_{4,5} + J_{4',5}$) for **2a**, **2b** and **2c** were determined as 15.9, 16.1 and 15.1 Hz, respectively.

^c In the case of aromatic protons *ortho*, *meta* and *para* are with respect to C-5.

^d The OCH₃ protons appeared at 3.89 ppm.

^e The *p*-Me protons appeared at 2.37 ppm.

^f The NMe₂ protons appeared at 2.89 ppm.

Table IV — ^{13}C NMR spectral data of cyclohex-2-enones **2a-h**

Compound	Chemical shifts (δ , ppm)								Aromatic ^a
	C-1	C-2	C-3	C-4	C-5	C-6	CH ₃ at C-3		
2a	199.1	126.8	161.7	39.0	40.8	43.9	24.4	143.3, 126.6	128.8, 127.0
2b	199.2	127.3	162.0	39.5	40.8	44.4	25.0	142.4, 128.8	129.9, 133.3
2c ^b	199.9	127.2	162.4	40.0	40.7	44.8	25.0	136.2, 128.3	114.8, 159.2
2d	198.0	127.1	161.2	38.6	40.9	43.6	24.7	147.4, 128.1	124.5, 150.9
2e ^c	199.7	126.9	162.3	39.5	40.8	44.9	24.8	140.7, 127.0	129.8, 136.9
2f	198.4	126.8	161.2	38.9	39.8	43.8	24.1	138.9, 128.1	115.0, 162.7
2g ^d	199.4	126.4	161.6	39.3	39.8	44.2	24.1	131.2, 128.2	112.8, 149.5
2h ^e	197.8	126.7	160.9	38.3	40.3	43.3	24.3	145.1, 122.1	133.0, 148.5 129.8, 121.7

^a Aromatic signals are mentioned in the order: *ipso*, *ortho*, *meta* and *para*.

^b OCH₃ signal was observed at 56.0 ppm.

^c *p*-Me signal was observed at 21.5 ppm.

^d NMe₂ signal was observed at 40.5 ppm.

^e Aromatic carbons are cited in the order *ipso*, *ortho*, *ortho'*, *meta*, *meta'* and *para*.

Tracey²², suggests that the torsional angle between H-5 and H-6' is about 175° and that between H-5 and H-6 is about 55°. The observed sum of the *vicinal* coupling constants about C(4)-C(5) bond also suggests that H-5 should be almost *anti* to one of the methylene protons at C-4 and *gauche* to the other methylene proton at C-4. These torsional angles strongly suggest that compounds **2a-h** adopt sofa conformation shown in **Figure 2**.

Comparison of the chemical shift of H-2 in **2a-h** with that in **5** suggests that the aryl group at C-5 deshields H-2 significantly. This is due to the magnetic field generated by the circulation of π -electrons in the aryl ring. The aryl group should adopt a conformation so that it is coplanar with C(5)-H(5) bond (**Figure 3**). In such a conformation H-2 should lie in the plane of the aryl group. Though H-2 is at a large distance from the center of the aryl group it should definitely lie in the deshielding region of the aryl group. It is also interesting to note that in **2a-h** H-5 is deshielded by about 1.2 ppm compared to that in **4**.

Comparison of the chemical shift of H-2 in **5** with that in **4** suggests that the 3-methyl group shields H-2 significantly. This is due to the magnetic anisotropic effect of C-CH₃ bond²³ (**Figure 4**). A similar shielding should be expected on the methylene protons at C-4. Hence, in **5** the average value of the chemical shifts of the methylene protons at C-4 should be less than that (2.36 ppm) in **4**. It is seen that in **2a-h** the average chemical shift of the methylene protons at C-4 is greater than that in **4**. Thus it is obvious that the aryl group at C-5 deshields the methylene protons at C-4 significantly. It is also obvious that the methylene protons at C-6 are deshielded by the aryl group at C-5.

The ¹³C chemical shifts of cyclohex-2-enone **4** (Ref., 21) are shown in **Figure 5**. Generally ¹³C chemical shifts are influenced by substituents in α and β -positions of the particular carbon²⁴. A γ -substituent causes a shielding if it is *gauche* to the concerned carbon. Also a highly electronegative substituent like F, OH or NH₂ causes a significant shielding though it is present in the γ -position *anti* to the concerned carbon²⁵.

Considering these factors it is obvious that the methyl group at C-3 should influence the chemical shifts of C-2, C-3 and C-4 significantly. Also the aryl group at C-5 should influence the chemical shifts of C-4, C-5 and C-6 significantly. Comparison of the ¹³C chemical shifts of **2a** and **4** suggests that the phenyl group deshields C-5 and C-6 by 18.2 and 5.9 ppm,

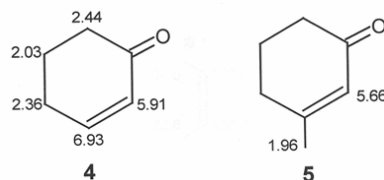


Figure 1—Proton chemical shifts of **4** and **5**

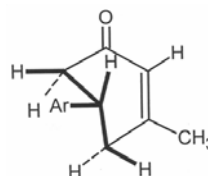


Figure 2—Conformation of cyclohex-2-enones **2a-2h**

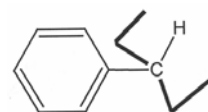


Figure 3—Conformation of phenyl group in **2a**

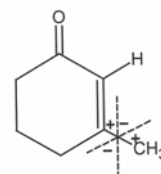


Figure 4—Shielding and deshielding regions of C-CH₃ bond

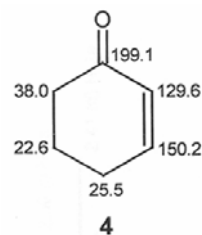


Figure 5—¹³C chemical shifts of cyclohex-2-enone **4** respectively. These effects are comparable to the α and β -effects of phenyl group in *trans*-4-*t*-butylphenyl-cyclohexane²⁶ which are 17.4 and 7.0 ppm, respectively. In **2a** C-4 is subjected to β -effect of phenyl and methyl groups. Comparison of the chemical shifts of C-4 in **2a** with that in **4** suggests that the combined β -effects of phenyl and methyl groups is 13.5 ppm. This is also comparable to the observed value of 12.8 ppm²⁶ in *t*-butylcyclohexanes. Comparison of chemical shifts of C-2 and C-3 in **3a** with those in **4** suggests that the CH₃ group deshields

C-3 (α -carbon) by 11.5 ppm and shields C-2 (β -carbon) by 2.7 ppm.

Cyanocyclohexanones **3a-c** and **3e-h**

Reaction of KCN and NH_4Cl with cyclic α,β -unsaturated ketones results in the formation of β -cyanoketones⁸. The reaction occurs by 1,4-addition followed by the transfer of a proton from oxygen to the α -carbon (**Scheme I**). Reaction of **2** with KCN and NH_4Cl can give rise to two stereoisomers **3** and **3'**. Since saturated six-membered ring compounds have been shown to exist largely in chair conformation the expected products should adopt conformation **3C** and **3'C**, respectively.

In the ^1H NMR spectrum of **3a** there was a triplet at δ 7.35 with $J = 7.4$ Hz, corresponding to two protons. This signal should be due to the *meta* protons of the phenyl group. There was a triplet at δ 7.26 with $J = 7.3$ Hz corresponding to one proton. This signal should be due to the *para* proton of the phenyl group. There was a doublet at δ 7.23 with $J = 7.2$ Hz corresponding to two protons. This signal should be due to the *ortho* protons of the phenyl group. In the observed spectrum the third line of the signal due to the *para* proton and the first line of the signal due to the *ortho* protons had overlapped.

There was a triplet of triplet at δ 3.36 corresponding to one proton. This signal should be due to the benzylic proton H-5. The benzylic proton is involved in two diaxial (*trans*) couplings viz., $J_{5a,6a}$ and $J_{4a,5a}$. It is also involved in two small *cis* couplings viz., $J_{5a,6e}$ and $J_{4e,5a}$. Sixteen lines with same intensity should be expected due to these couplings. However, the closely spaced lines overlap resulting in a triplet of triplet (**Figure 6**).

There was a singlet at δ 1.54, corresponding to three protons, due to the methyl protons. There were six one-proton signals in the range 1.9–2.7 ppm. These signals were assigned based on the observed correlations in the HOMOCOSY and NOESY spectra. The spectrum in this region along with the assignments is shown in **Figure 7**. The signal due to H-4a and H-6a appear as triplets instead of the expected double-doublets due to the overlap of the middle two closely spaced lines (**Figure 8**). This is again due to a small difference between J_{gem} and J_{aa} values. However, the spacing between the first and third lines of the observed triplets should give the sum of J_{gem} and J_{aa} . The signals for H-4e and H-6e must be

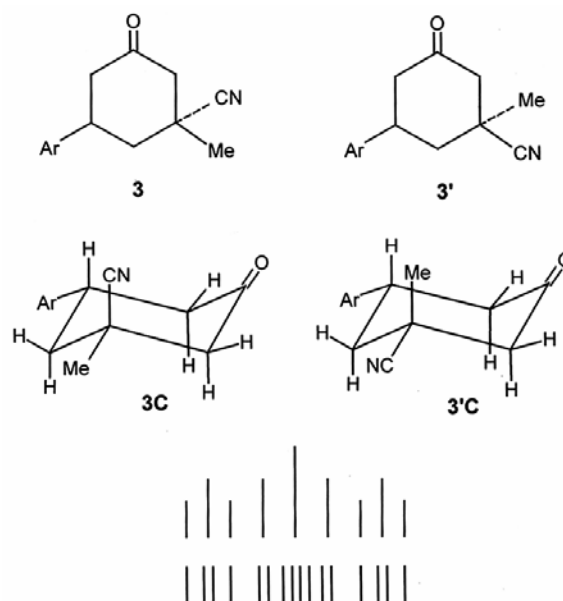


Figure 6—Overlap of lines for H-5

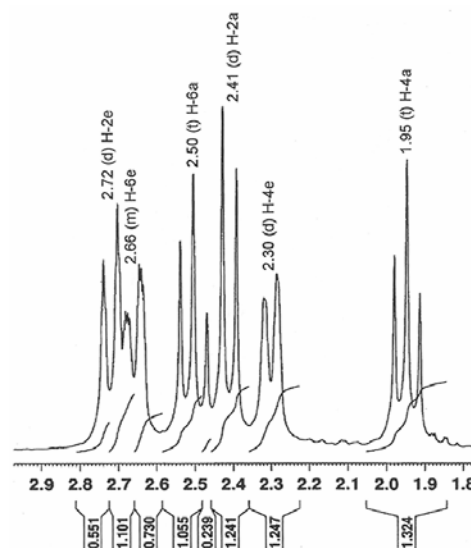


Figure 7—Proton NMR spectrum of **3a**



Figure 8—Overlap of lines for H-4a and H-6a

double-doublets. However, in the signal for H-6e additional splittings are seen. The signals for H-2e has four shoulders (each line of the doublet has two

shoulders). Though the signal for H-4e is not well resolved it also has same shoulders. These observations suggest that there is long range coupling among H-2e, H-4e and H-6e. This was also shown by the HOMOCOSY spectrum.

In the case of **3e** the signal due to H-6e contained

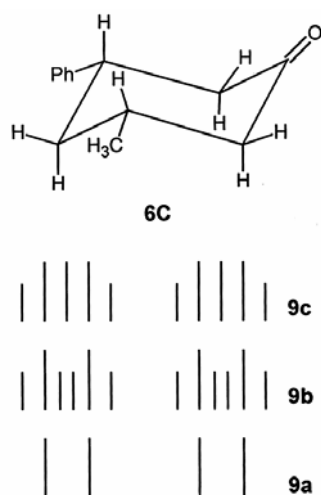


Figure 9—Overlap of lines for H-6e in **3e**

ten lines and that due to H-2e appeared as a triplet of doublet. This observation confirms the long range couplings among the three equatorial protons. The appearance of ten lines for H-6e can be explained as follows. H-6e undergoes coupling with H-6a and H-5a resulting in a doublet of doublet as shown in **Figure 9a**. Due to nearly equal long range couplings with H-4e and H-2e each line is split into a triplet as shown in **Figure 9b**. In the resulting six lines the middle two lines overlap resulting in the observation of 10 lines (**Figure 9c**). Indeed the intensities of lines 1, 5, 6 and 10 were about half of the intensities of the other six lines. Among these 10 lines, spacing between the first and sixth line should correspond to $J_{6a,6e}$. Indeed it was found that the spacings $\nu_1-\nu_6$, $\nu_2-\nu_7$, $\nu_3-\nu_8$, $\nu_4-\nu_9$ and $\nu_5-\nu_{10}$ had a constant value of 14.5 Hz. This value is $J_{6a,6e}$. The spacings $\nu_1-\nu_2$, $\nu_2-\nu_3$, $\nu_3-\nu_4$, $\nu_4-\nu_5$, $\nu_6-\nu_7$, $\nu_7-\nu_8$, $\nu_8-\nu_9$ and $\nu_9-\nu_{10}$ must correspond to the long range coupling. The average of these values was found as 1.9 Hz. Also the spacings $\nu_2-\nu_4$ and $\nu_7-\nu_9$ should correspond to $J_{6a,5a}$. Average of these values was found as 4.2 Hz.

In the case of **3a** six frequencies with almost same

Table V—Proton chemical shifts (δ , ppm) of cyanocyclohexanones **3a-c** and **3e-h**

Compound	Chemical shifts (δ , ppm)								Aromatic ^a
	H-2e	H-2a	H-4e	H-4a	H-5a	H-6e	H-6a	CH ₃	
3a	2.72	2.41	2.30	1.95	3.36	2.66	2.50	1.54	7.23, 7.35, 7.26
3b	2.78	2.37	2.36	1.95	3.41	2.72	2.49	1.57	7.24, 7.36
3c^b	2.75	2.38	2.31	1.89	3.35	2.67	2.45	1.56	7.16, 6.89
3e^c	2.76	2.46	2.34	1.88	3.37	2.70	2.45	1.55	7.16, 7.12
3f	2.76	2.41	2.33	1.92	3.38	2.68	2.47	1.57	7.21, 7.05
3g^d	2.75	2.36	2.31	1.87	3.32	2.68	2.45	1.55	7.12, 6.77
3h	2.83	2.42	2.40	2.00	3.57	2.77	2.52	1.61	8.15, 8.17, 7.57, 7.61

^a Chemical shifts are cited in the order *ortho*, *meta* and *para*.

^b The OCH₃ protons appeared at 3.80 ppm.

^c The *p*-Me protons appeared at 2.34 ppm.

^d The NMe₂ protons appeared at 2.95 ppm.

Table VI—Proton-proton coupling constants (J , Hz) in cyanocyclohexanones **3a-c** and **3e-h**

Compound	Coupling constants (J , Hz)							
	$J_{2a,2e}$	$J_{4a,4e}$	$J_{4a,5a}$	$J_{4e,5a}$	$J_{5a,6a}$	$J_{5a,6e}$	$J_{6a,6e}$	J_{ee}
3a	14.5	13.4	13.1	3.2	13.4	3.9	14.3	2.0
3b	14.5	13.4	13.2	3.1	13.2	4.1	14.5	2.0
3c	14.5	13.5	13.0	3.1	13.3	3.9	14.4	2.0
3e	14.5	14.5	13.0	3.5	13.3	3.9	14.5	1.9
3f	14.5	13.3	13.2	3.0	13.2	4.0	14.4	2.0
3g	14.5	13.7	12.8	3.1	13.3	4.0	14.4	2.0
3h	14.5	13.5	13.0	2.8	13.4	4.2	14.5	2.0

Table VII — ^{13}C chemical shifts (δ , ppm) of cyanocyclohexanones **3a-c** and **3e-h**

Compound	Chemical shifts (δ , ppm)									
	C-1	C-2	C-3	C-4	C-5	C-6	CH ₃	C≡N	Aromatic ^a	Others
3a	204.9	50.3	36.2	42.7	40.5	47.1	26.8	122.1	141.8, 126.3, 128.6, 127.6	
3b	204.3	50.6	36.3	43.1	40.1	47.3	27.3	122.1	140.5, 127.9, 129.1, 133.0	
3c	205.3	50.3	36.1	43.0	39.8	47.4	26.9	122.2	134.0, 127.3, 114.0, 158.4	55.0 (OCH ₃)
3e	205.0	51.0	36.6	43.8	40.6	47.8	27.7	122.0	139.4, 126.6, 129.7, 137.3	21.2 (CH ₃)
3f	205.1	50.6	36.5	43.2	40.2	47.6	27.2	122.5	138.1, 128.3, 115.8 (d, J = 21.0 Hz) 161.8 (d, J = 244 Hz)	
3g	205.6	50.8	36.5	43.8	40.1	47.9	27.4	122.6	130.6, 127.3, 113.3, 149.6	40.9 N(CH ₃) ₂
3h^b	203.6	50.7	36.4	42.9	40.4	47.1	27.4	121.8	144.1, 121.3, 138.5, 148.8, 126.8, 122.0	

^aAromatic signals are cited in the order *ipso*, *ortho*, *meta*, *para*.^bAromatic carbons are cited in the order *ipso*, *ortho*, *ortho'*, *meta*, *meta'* and *para*.

intensity were given by the computer. Obviously, in this case the lines 1, 5, 6 and 10 with half intensity of the other lines have not appeared clearly. This may be due to the recording of the ^1H NMR spectrum of **3e** at 500 MHz and that of **3a** at 400 MHz. However, even in this case the values of $J_{5a,6a}$, $J_{6a,6e}$ and long range coupling could be computed from the observed six frequencies.

From the frequencies of the three lines for H-6a the value of $J_{5a,6a}$ could be computed since the value of $J_{6a,6e}$ is known. From the data for H-4e and H-4a the values of $J_{4a,4e}$ and $J_{4a,5a}$ could be computed.

Though $J_{2e,4e}$, $J_{4e,6e}$ and $J_{6e,2e}$ may be expected to be nearly equal the values $J_{5a,6e}$ and $J_{5a,4e}$ may differ significantly. Hence, in the signal for H-5 the spacing between the first and second lines may not correspond to either $J_{5a,4e}$ or $J_{5a,6e}$. However, the spacings ν_2 - ν_3 , ν_5 - ν_6 and ν_7 - ν_9 of this signal should give the sum of $J_{5a,4e}$ and $J_{5a,6e}$. From the known value of $J_{5a,6e}$ the value of $J_{5a,4e}$ could be computed. Such calculations could be made for all compounds except **3b**.

The ^{13}C signals of **3a** could be assigned based on positions, intensities, SEFT spectrum and the correlations in the HSQC spectrum. The ^{13}C signals for the other cyanocyclohexanones were assigned by comparison with **3a**. In **3b**, **3c** and **3e-3h** the aromatic ^1H and ^{13}C signals were assigned based on known substituent effects^{19,20}. The various proton chemical shifts of the cyanocyclohexanones are given in **Table V**. The various proton-proton coupling constants are given in **Table VI**. The ^{13}C chemical shifts are given in **Table VII**.

From the NOESY spectrum of **3a** it was inferred that the methyl protons have strong NOE with H-2a, H-2e, H-6a and H-6e. However, there was no NOE between the methyl protons and H-5. From the observed vicinal coupling constants and NOEs it is obvious that compound **3a** should exist in conformation **3aC** (**3C** with Ar = C₆H₅). Indeed from the X-ray crystallographic study²⁷, of **3a** it has been found that **3a** exists in chair conformation **3aC**. Also X-ray crystallographic study²⁸ of **3h** has shown that it adopts chair conformation **3hC** (**3C** with Ar = *m*-NO₂C₆H₄).

It has been found²⁹ that an equatorial methyl group shields an adjacent axial proton by 0.42 to 0.45 ppm, whereas an axial methyl group deshields it by 0.16-0.26 ppm. The effect of axial and equatorial cyano groups on the chemical shifts of various protons in cyclohexane have been determined by Abraham and Reid³⁰. Based on the effects of the methyl and cyano groups H-2a and H-4a in **3'C** should be deshielded by 0.59-0.72 ppm than those in **3C**. However, in **3b**, **3c**, **3e**, **3f** and **3g** the chemical shifts of H-2a and H-4a differ from those in **3a** only by 0.00-0.08 ppm. Hence, it is obvious that all these compounds should adopt conformation **3C**.

It is of interest to find the effect of the double bond at C-2 on the chemical shifts of the saturated carbons C-4, C-5 and C-6 and the protons on these carbons. This can be done by comparing the chemical shifts of appropriate carbons and protons in 3*r*-methyl-5*c*-phenylcyclohexanone **6** with the corresponding carbons and protons in **2a**. Compound **6** should adopt conformation **6C** with equatorial orientations of the phenyl and methyl groups.

Effect of axial cyano group on all protons in **3a** are available³⁰ except for H-6e. However, the axial cyano group at C-3 can influence the chemical shifts of protons at C-6 only by magnetic anisotropic effect which depends on the relative orientations of the cyano group and the particular proton. The effect of axial cyano group at C-3 on H-6e may be taken as that of an equatorial cyano group at C-3 on H-6a, for which the value is available. Using the SCS values for cyano group and from the chemical shifts of **3a** the probable values for the chemical shifts of H-4e, H-4a, H-5a, H-6e and H-6a in **6** were computed as 2.01, 1.62, 2.90, 2.58 and 2.45 ppm, respectively. Comparison of these values with those of the corresponding protons (H-4, H-4'), H-5, H-6 and H-6' in **2a** suggests that the double bond at C-2 deshields all these protons significantly.

From the known effects of phenyl and methyl group²⁴ on ¹³C chemical shifts and the chemical shifts of carbon of cyclohexanone³¹ the probable chemical shifts of C-4, C-5 and C-6 in **6** were computed as 40.0, 44.0 and 48.9 ppm, respectively. In these calculations the small γ -anti effects of phenyl and methyl groups were ignored. Comparison of these values with those in **2a** suggests that these carbons are shielded by the double bond at C-2.

It is seen that H-5 is deshielded by 0.44 ppm and C-5 is shielded by 3.2 ppm by the double bond at C-2. This may be due to a polarization of C(5)-H(5) bond by the π -electron cloud so that H-5 gets a slight positive charge and C-5 gets a slight negative charge. Similar effects by an axial hydroxy group and ¹H and ¹³C chemical shifts have been found as 0.49²⁹ and 6.7 ppm²⁴, respectively. Thus, the observed deshielding on H-5 is not proportional to the observed shielding on C-5. Probably, the deshielding on H-5 may be partly due to magnetic anisotropy of the C=C bond.

Also H-6 and H-6' are shielded by 0.08 and 0.13 ppm, respectively whereas C-6 is shielded by 5 ppm by the double bond at C-2. These effects may be due to a combination of polarization of C-(H-6') bond, magnetic anisotropic effect and a decrease in the electronegativity of the carbonyl group. The contribution of structure **C** to the resonance should decrease the positive charge on the carbonyl carbon which in turn should decrease the electronegativity of the carbonyl group.

It is seen that H-4 (equatorial like proton) and H-4' (axial like proton) are deshielded by 0.46 and 0.75

ppm, respectively, by the double bond at C-2. However, C-4 is shielded only by 1.0 ppm by the double bond at C-2. This is in contrast to the observations made on H-6, H-6' and C-6.

In the cyclohexanone **6** C-3 is a saturated carbon whereas in **2b** it is an olefinic carbon. Furthermore, due to contribution of structure **C** to the resonance the electronegativity of C-3 is enhanced significantly. This increase in electronegativity should deshield H-4, H-4' and C-4. The polarisation of C-(H-4') bond shields C-4 but deshields H-4'. Thus, the two effects act in opposite directions for C-4 but in the same direction for H-4'. Hence, H-4' is deshielded significantly but there is a small net shielding on C-4.

Experimental Section

Preparation of compounds

3*t*-Aryl-2*r*,4*c*-dicarbalkoxy-5*c*-hydroxy-5*t*-methylcyclohexanones **1**

These compounds were prepared following the methods already reported¹⁵⁻¹⁷. All compounds were purified by recrystallization from ethanol. The melting points were determined and there was good agreement with these reported already. Compounds **2e** and **3e** were prepared starting from 3*t*-(*p*-tolyl)-2*r*,4*c*-bis(isopropoxycarbonyl)-5*c*-hydroxy-5*t*-methylcyclohexanone for which the physical and spectral data have not been reported earlier. These are as follows: Yield 80%; m.p. 164 °C. IR: 3483 (for OH); 1735, 1709 cm⁻¹ (for C=O). ¹H NMR (500 MHz, CDCl₃): δ 0.61, 1.01, 1.05, 1.10 (doublets each corresponding to 3H, CH₃ of COOCH(CH₃)₂ at C-2 and C-4); 1.32 (s, 3H, CH₃ at C-5); 2.28 (s, 3H, CH₃ at phenyl); 2.46 (dd, 1H, *J* = 3.00, 14.5 Hz, H-6a); 2.68 (d, 1H, *J* = 14.5 Hz, H-6e); 2.96 (d, 1H, *J* = 12.0 Hz, H-4a); 3.58 (d, 1H, *J* = 12.5 Hz, H-2a); 3.75 (d, *J* = 2.5 Hz, OH at C-5); 3.93 (t, 1H, *J* = 12.3 Hz, H-3a); 4.77, 4.80 (multiplets each corresponding to 1H, CH of COOCH(CH₃)₂ at C-2 and C-4); 7.26, 7.09 (aromatic protons). ¹³C NMR (125 MHz, CDCl₃): 20.9, 21.7 (CH₃ of COOCH(CH₃)₂ at C-2 and C-4); 21.7 (CH₃ of *p*-tolyl); 28.8 (CH₃ at C-5); 45.1 (C-3); 53.0 (C-6); 57.4 (C-4); 63.0 (C-2); 68.7, 68.9 (CH of COOCH(CH₃)₂ at C-2 and C-4); 73.2 (C-5); 128.3, 129.3 (aromatic carbons); 135.3, 137.5 (*ipso* carbons); 167.4, 173.6 (CO of COOCH(CH₃)₂ at C-2 and C-4); 201.5 (C=O).

5-Aryl-3-methylcyclohex-2-enones **2a-h**

A mixture of the appropriate ketone **1** (0.01 mole) in glacial acetic acid (25 mL) and concentrated

hydrochloric acid (50 mL) was refluxed for 9 hr. After completion of the reaction, the reaction-mixture was neutralized with ammonia and the product was purified by column chromatography using silica gel (100–200 mesh) with benzene-ethyl acetate (9.5:0.5) as eluent.

5*r*-Aryl-3*t*-cyano-3*c*-methylcyclohexanones (3*a*-*c* and 3*e*-*h*)

A mixture of 5-aryl-3-methylcyclohex-2-enones (0.02 mole), potassium cyanide (0.04 mole), ammonium chloride (0.03 mole), dimethylformamide (50 mL) and water (2 mL) was heated and stirred for about 18–20 hr at 80°C. During this time ammonia gas was evolved. The reaction mixture was poured into water and extracted with CH₂Cl₂ (30 mL). The organic layer was washed with water and dried over anhydrous sodium sulphate. Evaporation of solvent gave the crude product which was purified by column chromatography using silica gel (100–200 mesh) as adsorbent and hexane-ethyl acetate (4.5:1) as eluent.

Recording of spectra

¹H NMR spectra

For **2a**, **2d** and **2e** ¹H NMR spectra were recorded on a Bruker AV 300 NMR spectrometer operating at 300.12 MHz for ¹H with the following parameters: number of scans 32; data points 32 K; sweep width 6172.84 Hz; acquisition time 2.65 sec. For **2b**, **2c** and **2h** ¹H NMR spectra were recorded on a Jeol GSX 400 NMR spectrometer operating at 399.65 MHz for ¹H with the following parameters: number of scans 64; data points 32 K; sweep width 5000 Hz; acquisition time 2.05 sec. The ¹H NMR spectrum of **3e** was recorded on a Bruker DRX 500 NMR spectrometer operating at 500.03 MHz for ¹H with the following parameters: number of scans 284; data points 32 K; sweep width 34013.61 Hz; acquisition time 0.48 sec. For all the other compounds ¹H NMR spectra were recorded on a Bruker AMX 400 NMR spectrometer operating at 400.14 MHz for ¹H with the following parameters: number of scans 64; data points 16 K; sweep width 7246.38 Hz; acquisition time 1.26 sec. For recording ¹H NMR spectra, solutions were prepared by dissolving 10 mg of the material in 0.5 mL of CDCl₃.

¹³C NMR spectra

For **2d**, **2e** and **2h** ¹³C NMR spectra were recorded on a AV 300 NMR spectrometer operating at 75.48

MHz for ¹³C and with the following parameters: number of scans 400; data points 64 K; sweep width 17985.61 Hz; acquisition time 1.82 sec. For **2b**, **2c** and **3e** ¹³C NMR spectra were recorded on a Bruker DRX 500 NMR spectrometer operating at 125.77 MHz for ¹³C with the following parameters: number of scans 284; data points 16 K; sweep width 31446.54 Hz; acquisition time 0.26 sec. For other compounds ¹³C NMR spectra were recorded on a Bruker AMX 400 NMR spectrometer operating at 100.63 MHz for ¹³C with the following parameters: number of scans 256; data points 16 K; sweep width 31446.54 Hz; acquisition time 0.26 sec. For recording ¹³C NMR spectra, solutions were prepared by dissolving 50 mg of the material in 0.5 mL of CDCl₃.

2D NMR spectra

For **3a**, HOMOCOSY, HMBC, HSQC spectra were recorded on a Bruker DRX 500 NMR spectrometer and for **2e**, HMBC and HSQC spectra were recorded on a Bruker AV 300 NMR spectrometer using standard parameters. Phase sensitive NOESY spectrum of **3a** was recorded with a mixing time 1 sec.

For recording 2D NMR spectra, solutions were made by dissolving 50 mg of the material in 0.5 mL of the solvent. The number of data points was 1 K. All NMR measurements were made using 5 mm tubes.

IR spectra recorded on Avatar 330 FT-IR Thermo Nicolet spectrometer in KBr pellets. The mass spectrum of **3a** was recorded on a Jeol DX-303 mass spectrometer. Elemental analyses were performed on a Perkin-Elmer CHNS/O analyser.

Conclusion

The observed *vicinal* coupling constants of **2a-h** suggest that these compounds adopt a sofa conformation in which C-1, C-2, C-3, C-4, C-6 lie almost in one plane and C-5 lies above this plane. The NMR spectral data of the cyanocyclohexanones suggest that these compounds exist in chair conformation with cyano group in the axial position and the aryl and methyl groups in the equatorial positions. Comparison of chemical shifts of **2a** with the probable value of 3*r*-methyl-5*c*-phenylcyclohexenone suggests that the double bond at C-2 influences the chemical shifts of C-4, C-5, C-6 and the protons on these carbons by polarisation, magnetic anisotropy and electronegativity effects.

Acknowledgement

Thanks are due to NMR Research Centre, IISc, Bangalore for recording the NMR spectra.

References

- 1 Hernandez R & Suarez E, *J Org Chem*, 59, **1994**, 2766.
- 2 Cocker W, Grayson D H & Shannon P V R, *J Chem Soc Perkin Trans I*, **1995**, 1153.
- 3 Sobolev A, Vos M, Zuilhof H T, Saraber F C E, Jansen B J M & de Groot Ae, *ARKIVOC*, **2005**, 29.
- 4 Lewandowska E, *Tetrahedron*, 63, **2007**, 2107.
- 5 Rodig O R & Johnston N J, *J Org Chem*, 34, **1969**, 1949.
- 6 Nagata W, Hirai S, Itazaki H & Takeda K, *J Am Chem Soc*, 26, **1961**, 2413.
- 7 Meyer W L & Wolfe J F, *J Org Chem*, 29, **1964**, 170.
- 8 Pandey S C, Singh S S, Patro B & Ghosh A C, *Indian J Chem*, 43B, **2004**, 2705.
- 9 Hawkes G E, Herwig K, & Roberts J D, *J Org Chem*, 20, **1974**, 1017.
- 10 Ramalingam K, Berlin K D, Loghry R A, Van der Halm D & Satyamurthy N, *J Org Chem*, 44, **1979**, 477.
- 11 Pandiarajan K, Sabapathy Mohan R T & Hasan M U, *Magn Reson Chem*, 24, **1986**, 312.
- 12 Pandiarajan K, Sabapathy Mohan R T & Krishnakumar B, *Indian J Chem*, 26B, **1987**, 624.
- 13 Caldwell C W, Gawhie A D, Mills J N & Greco M, *Magn Reson Chem*, 31, **1993**, 309.
- 14 Gurumani V, Pandiarajan K & Swaminathan M, *Bull Chem Soc Japan*, 70, **1997**, 29.
- 15 Pandiarajan K, Sabapathy Mohan R T, Gomathi R & Muthukumaran G, *Magn Reson Chem*, 43, **2005**, 430.
- 16 Amirthaganesan S, Sabapathy Mohan R T, Murugavel K, Muthukumaran G & Pandiarajan K, *Indian J Chem*, 46B, **2007**, 1004.
- 17 Pandiarajan K, Sabapathy Mohan R T, Murugavel K & Hema R, *J Mol Strut*, 875, **2008**, 226.
- 18 Sivasubramanian S, Ponnuswamy A & Muthusubramanian S, *Indian J Chem*, 34B, **1995**, 825.
- 19 Silverstein R M & Webster F X, *Spectrometric Identification of Organic Compounds*, 6th edn. (Wiley; New York), **2007**, p 209.
- 20 Silverstein R M & Webster F X, *Spectrometric Identification of Organic Compounds*, 6th edn. (Wiley; New York), **2007**, p 229.
- 21 Reich H J, *Proton and Carbon-13 Chemical Shifts of Cyclohex-2-enone and 3-Methylcyclohex-2-enone*, (University of Wisconsin: Madison), **2008**.
- 22 Slessor K N & Tracey A S, *Can J Chem*, 49, **1971**, 2874.
- 23 Silverstein R M & Webster F X, *Spectrometric Identification of Organic Compounds*, 6th edn. (Wiley; New York), **1998**, p 155.
- 24 Pandiarajan K & Manikekalai A, *Magn Reson Chem*, 31, **1993**, 80.
- 25 Eliel E L, Bailey W F, Kopp L D, Wiler R L, Grant D M, Bertrand R, Christensen K A, Dalling D K, Duch M W, Wenkert E, Schell P M & Gochran D W, *J Am Chem Soc*, 97, **1975**, 322.
- 26 Eliel E L & Manoharan M, *J Org Chem*, 46, **1981**, 1959.
- 27 Subramanyam M, Thiruvalluvar A, Sabapathy Mohan R T & Kamatchi S, *Acta Crystallogr*, 63E, **2007**, 2715.
- 28 Subramanyam M, Thiruvalluvar A, Sabapathy Mohan R T & Kamatchi S, *Acta Crystallogr*, 63E, **2007**, 2717.
- 29 Pandiarajan K, Manimekalai A & Rajarajan G, *Indian J Chem* 39B, **2000**, 517.
- 30 Abraham R J & Reid M, *Magn Reson Chem*, 38, **2000**, 570.
- 31 Silverstein R M & Webster F X, *Spectrometric Identification of Organic Compounds*, 6th edn. (Wiley; New York), **2007**, p 234.